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Term: 19 with 16 with vector 

Display: 10 **Documents in Display Format:** - **Starting with Number** 1

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Search History

DATE: Monday, February 16, 2004 [Printable Copy](#) [Create Case](#)

<u>Set Name</u> side by side	<u>Query</u>	<u>Hit Count</u>	<u>Set Name</u> result set
<i>DB=PGPB,USPT,USOC,EPAB,JPAB,DWPI; PLUR=YES; OP=ADJ</i>			
<u>L30</u>	19 with 16 with vector	7	<u>L30</u>
<u>L29</u>	115 with vector	18	<u>L29</u>
<u>L28</u>	L27 same 16	0	<u>L28</u>
<u>L27</u>	produc\$ with 12	1978	<u>L27</u>
<u>L26</u>	L25 same 16	0	<u>L26</u>
<u>L25</u>	packagi\$ with 12	1492	<u>L25</u>
<u>L24</u>	L23 same 122	0	<u>L24</u>
<u>L23</u>	replication defective	5569	<u>L23</u>
<u>L22</u>	119 same 16	118	<u>L22</u>
<u>L21</u>	119 same 117	0	<u>L21</u>
<u>L20</u>	L19 same 14	7779	<u>L20</u>
<u>L19</u>	aggregation	63508	<u>L19</u>
<u>L18</u>	L17 same filtra\$	0	<u>L18</u>
<u>L17</u>	16 same 12	44	<u>L17</u>
<u>L16</u>	L15 same 12	2	<u>L16</u>
<u>L15</u>	L14 with 16	295	<u>L15</u>

<u>L14</u>	improv\$ or enhanc\$	5308508	<u>L14</u>
<u>L13</u>	L12 same l2	11	<u>L13</u>
<u>L12</u>	l9 same l6	2239	<u>L12</u>
<u>L11</u>	L10 with l6	3	<u>L11</u>
<u>L10</u>	L9 with l2	345	<u>L10</u>
<u>L9</u>	stabil\$	1738553	<u>L9</u>
<u>L8</u>	L7 same l4	15	<u>L8</u>
<u>L7</u>	L6 same l2	44	<u>L7</u>
<u>L6</u>	human serum albumin	10793	<u>L6</u>
<u>L5</u>	L4 same l3	7	<u>L5</u>
<u>L4</u>	concentration or titer	1296548	<u>L4</u>
<u>L3</u>	L2 with l1	60	<u>L3</u>
<u>L2</u>	adenovir\$	29148	<u>L2</u>
<u>L1</u>	HSa or human serum albumin	13009	<u>L1</u>

END OF SEARCH HISTORY

First Hit**End of Result Set**☐ **Generate Collection** **Print**

L5: Entry 7 of 7

File: DWPI

Jul 1, 2003

DERWENT-ACC-NO: 2000-206000

DERWENT-WEEK: 200366

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TITLE: New composition useful for preservation of viral particles by enhancing vector titer and/or stabilizing vector at refrigerator or room temperature, comprising recombinant adenovirus vector and human serum albumin

INVENTOR: MCGLENNON, K R; MOODY, D ; SHIH, S

PATENT-ASSIGNEE: AVENTIS PHARM INC (AVET), AVENTIS PHARM PROD INC (AVET)

PRIORITY-DATA: 1998US-096600P (August 14, 1998)

Search Selected**Search ALL****Clear**

PATENT-FAMILY:

PUB-NO	PUB-DATE	LANGUAGE	PAGES	MAIN-IPC
<input type="checkbox"/> <u>MX 2001001727 A1</u>	July 1, 2003		000	A61K035/00
<input type="checkbox"/> <u>WO 200009675 A1</u>	February 24, 2000	E	053	C12N015/00
<input type="checkbox"/> <u>AU 9954858 A</u>	March 6, 2000		000	C12N015/00
<input type="checkbox"/> <u>EP 1109896 A1</u>	June 27, 2001	E	000	C12N015/00
<input type="checkbox"/> <u>AU 748523 B</u>	June 6, 2002		000	C12N015/00
<input type="checkbox"/> <u>JP 2003528029 W</u>	September 24, 2003		064	A61K048/00

DESIGNATED-STATES: AE AL AM AT AU AZ BA BB BG BR BY CA CH CN CU CZ DE DK EE ES FI GB GD GE GH GM HR HU ID IL IN IS JP KE KG KP KR KZ LC LK LR LS LT LU LV MD MG MK MN MW MX NO NZ PL PT RO RU SD SE SG SI SK SL TJ TM TR TT UA UG US UZ VN YU ZA ZW AT BE CH CY DE DK EA ES FI FR GB GH GM GR IE IT KE LS LU MC MW NL OA PT SD SE SL SZ UG ZW AL AT BE CH CY DE DK ES FI FR GB GR IE IT LI LT LU LV MC MK NL PT RO SE SI

APPLICATION-DATA:

PUB-NO	APPL-DATE	APPL-NO	DESCRIPTOR
MX2001001727A1	August 13, 1999	1999WO-US18515	
MX2001001727A1	February 14, 2001	2001MX-0001727	
MX2001001727A1		WO 200009675	Based on
WO 200009675A1	August 13, 1999	1999WO-US18515	
AU 9954858A	August 13, 1999	1999AU-0054858	
AU 9954858A		WO 200009675	Based on
EP 1109896A1	August 13, 1999	1999EP-0941147	
EP 1109896A1	August 13, 1999	1999WO-US18515	

EP 1109896A1		WO 200009675	Based on
AU 748523B	August 13, 1999	1999AU-0054858	
AU 748523B		AU 9954858	Previous Publ.
AU 748523B		WO 200009675	Based on
JP2003528029W	August 13, 1999	1999WO-US18515	
JP2003528029W	August 13, 1999	2000JP-0565112	
JP2003528029W		WO 200009675	Based on

INT-CL (IPC): A61 K 35/00; A61 K 35/76; A61 K 38/00; A61 K 47/10; A61 K 47/26; A61 K 48/00; A61 P 9/00; A61 P 25/00; A61 P 35/00; C07 H 21/04; C12 N 15/00; C12 N 15/09; C12 N 15/63

ABSTRACTED-PUB-NO: WO 200009675A

BASIC-ABSTRACT:

NOVELTY - A composition (I), comprising a recombinant adenovirus vector (II) and a concentration of human serum albumin (HSA), is new and stabilizes (II) at a temperature above the freezing point of water or enhances a titer of (II) compared to a titer in the absence of HSA, or both, in an aqueous buffer.

DETAILED DESCRIPTION - INDEPENDENT CLAIMS are also included for the following:

(1) a method (III) for preparing a stabilized recombinant adenovirus formulation; and

(2) a method (IV) for stabilizing an adenovirus vector at about 20 deg. C, comprising preparing an admixture of the adenovirus vector in an aqueous composition of Dulbecco's phosphate buffered saline, from about 5 - 15% glycerol, from 0.25 - 2.0 mM CaCl₂ and from 0.1 - 1.0 mM MgCl₂.

USE - (I) and (IV) are useful for the preservation and/or storage of viral particles and viral vectors which is directly injectable into an organism, especially for gene therapy in human and veterinary medicine.

ADVANTAGE - (I) optimally enhances the vector titer and/or stabilizes the vector at refrigerator or room temperature. The solution is available for administration immediately after removal from the storage temperature, without any further manipulation. It is also possible to carry out the removal from storage conditions directly in the clinic therefore reducing time between storage and use which also makes it possible to remain constantly in sterile formulation and therefore reduce the risks of external contamination.

ABSTRACTED-PUB-NO: WO 200009675A

EQUIVALENT-ABSTRACTS:

CHOSEN-DRAWING: Dwg.0/11

DERWENT-CLASS: B04 C06 D16

CPI-CODES: B04-B04D2; B04-E08; B04-F1100E; B04-H0100E; B04-N04; B14-S03; C04-B04D2; C04-E08; C04-H0100E; C04-N04; C14-S03; D05-C07; D05-H12E; D05-H17;

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L3: Entry 17 of 60

File: PGPB

Aug 29, 2002

DOCUMENT-IDENTIFIER: US 20020119942 A1

TITLE: Packaging systems for human recombinant adenovirus to be used in gene therapy

Detail Description Paragraph:

[0245] In a non-limiting example we describe the generation and functionality of a recombinant adenovirus containing the murine HSA gene in the E1 region and the firefly luciferase gene in the gp19K region. The luciferase gene was excised from pAd/MLP-Luc (described in EP 0707071) as a Hind III-BamH I construct and cloned into the Hind III-BamH I sites of pBS.Eco-Eco/ad5.DELTA.HIII.DELTA.gp19K.DELTA.XbaI. Then the Msc I-Mun I fragment containing the luciferase gene was cloned into the corresponding sites of pBS.Eco-Eco/ad5.DELTA.gp19K generating pBS.Eco-Eco/ad5.DELTA.gp1-9K.luc. This restores the Eco-Eco fragment, but now with the luciferase gene in the place of gp 19K.

Detail Description Paragraph:

[0248] The construct pAd5/S430-HSA also was digested with Xba I and Sca I and the 1252 bp fragment (containing the remainder of the ampicillin gene, the left ITR and packaging signal from adenovirus and the 5' part of the S430 promoter) was isolated. A third fragment of 1576 bp was isolated from the MFG-S-based retroviral vector following an XbaI digestion and contains MFG-S sequences corresponding to bp 695-2271.

Detail Description Paragraph:

[0287] A minimal adenoviral vector contains (as operably linked components) the adenovirus-derived cis elements necessary for replication and packaging, with or without foreign nucleic acid molecules to be transferred. Recently, the lower limit for efficient packaging of adenoviral vectors has been determined at 75% of the genome length (Parks and Graham, J. Virol. 71(4):3293-8 (1997)). To allow flexible incorporation of various lengths of stuffer fragments, a multiple cloning site (MCS) was introduced into a minimal adenoviral vector. To obtain a minimal adenoviral vector according to the invention, the following constructs were made: pAd/L420-HSA (FIG. 21) was digested with Bgl II and Sal I and the vector-containing fragment was isolated. This fragment contains the left ITR and packaging signal from Ad5 and the murine HSA gene driven by a modified retroviral LTR. The right ITR of adenovirus was amplified by PCR on pBr/Ad.BamH I-rITR template DNA using the following primers: PolyL-ITR: 5'-AAC-TGC-AGA-TCT-ATC-GAT-ACT-AGT-CAA-TTG-CTC-GAG-TC-T-AGA-CTA-CGT-CAC-CCG-CCC-CGT-TCC-3' (SEQ. ID. NO. 32) and ITR-BSN: 5'-CGG-GAT-CCG-TCG-ACG-CGG-CCG-CAT-CAT-CAA-TAA-TAT-ACC-3' (SEQ. ID. NO. 33). The amplified fragment was digested with Pst I and BamHI and cloned into pUC 119 digested with the same enzymes. After sequence confirmation of correct amplification of the ITR and the MCS, a Bgl II-Sal I fragment was isolated and cloned into the Bgl II-Sal I digested pAd/L420-HSA fragment described above. The resulting clone was named pAd/L420-HSA.ITR.

CLAIMS:

7. The recombinant nucleic acid of claim 6, wherein said adapter plasmid is pAd5/CLIP or pAd5/L420-HSA, and said recombinant nucleic acid comprises adenovirus derived nucleotides 1-454; and adenovirus nucleotides 3511-6095.

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L3: Entry 18 of 60

File: PGPB

Aug 29, 2002

DOCUMENT-IDENTIFIER: US 20020117170 A1

TITLE: Compositions and methods for the pulmonary delivery of aerosolized macromolecules

Detail Description Paragraph:

[0196] Bulk pCMV.beta. DNA:Adenovirous vector as described in U.S. application Ser. No: 08/417,507 filed Apr. 14, 1995 entitled, "COMPOSITIONS AND METHODS FOR NUCLEIC ACID DELIVERY TO THE LUNG", the disclosure of which is hereby incorporated by reference, was obtained from Genzyme Corporation, Cambridge, Mass. A DNA:adenovirous vector formulation was achieved by combining 108 PFU/mL DNA:Lipid vector per 1.0 mL deionized water with 6.1 mg/mL glycine J. T. Baker) 2.5 mg/mL HSA, 1.9 mg/mL phosphate buffer at pH 7.4.

Detail Description Paragraph:

[0202] The above DNA:adenovirous vector dry powder composition contained 58% glycine, and 24% HSA and 18% phosphate buffer.

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L3: Entry 30 of 60

File: USPT

Jun 24, 2003

DOCUMENT-IDENTIFIER: US 6582729 B1

**** See image for Certificate of Correction ****

TITLE: Powered pharmaceutical formulations having improved dispersibility

Detailed Description Text (102):

This example includes two sets of experiments. (A) In the first set, we investigated the effects of carbohydrate and amino acid excipients in phosphate buffer (PB), (i) Mannitol/HSA, (ii) Glycine/HSA and (iii) Mannitol/Glycine/HSA, on the infectivity of the adenovirus dry powders. (B) In the second set, we investigated the effects of buffer removal and the process outlet temperature on the infectivity. All solutions were used and stored cold (.about.5.degree. C.).

Detailed Description Text (103):

(A) (i) To 4.times.3 ml Mannitol/HSA it was added 0.1 ml of adenovirus solution to obtain 3.2.times.10.sup.7 iu/ml and .about.60 mg/ml solids, and the fifth was used as a control with no virus. Two of the virus formula were diluted with de-ionized water to .about.9 mg/ml solids. (ii) Two formulations of 6.3 ml Glycine/HSA (I) in PB plus 0.4 ml adenovirus solution were made (29 mg/ml solids, 6.3.times.10.sup.7 iu/ml). One of them was diluted with de-ionized water to 9 mg/ml solids. (iii) Two formulations of 4.1 ml Mannitol/Glycine/HSA in PB plus 0.4 ml of virus solution were made (45.1 mg/ml solids, 8.89.times.10.sup.7 iu/ml). One was diluted with de-ionized water to 9 mg/ml. The adenovirus solution was freshly made on the same day and was kept cold on ice.

Detailed Description Text (104):

(B) Four formulations were prepared wherein two contained 25 ml of Glycine/HSA (II) in PB plus 0.4 ml of adenovirus solution (10.5 mg/ml, 1.6.times.10.sup.7 iu/ml) and the other two contained 25 ml of Glycine/HSA (II) in water plus 0.4 ml of adenovirus solution (8.6 mg/ml, 1.6.times.10.sup.7 iu/ml). The adenovirus solution underwent only one freeze/thaw cycle before usage in the above preparations. It was prepared about 10 weeks prior and was stored frozen at -70.degree. C.

Detailed Description Paragraph Table (4):

TABLE 3 Glycine/HSA Adenovirus Formulations Formula Dipersi. HORIBA Cascade impactor % infectivity (mg/ml) (% RSD) MMD MMAD % < 5 .mu.m retained 29 40(25) 2.6 2.8 70 14 9 51(1) 2.3 1.8 80 7

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L3: Entry 44 of 60

File: USPT

Oct 16, 2001

DOCUMENT-IDENTIFIER: US 6303582 B1

TITLE: Compositions and methods for nucleic acid delivery to the lung

Detailed Description Text (118):

This study included two sets of experiments. In the first set, the effects of bulking agents in phosphate buffer (PB), (i) mannitol/HSA, (ii) glycine/HSA and (iii) mannitol/glycine/HSA, on the infectivity of adenovirus dry powders were investigated. In the second set, the effects of buffer removal and process outlet temperature on viral infectivity were investigated. All solutions were used and stored cold (about 5.degree. C.).

Detailed Description Text (120):

(i) Five mannitol/HSA in PB formulations were prepared as follows: To four samples of 4.times.3 ml mannitol/HSA was added 0.1 ml of adenovirus solution to obtain 3.2.times.10.sup.7 iu/ml and about 60 mg/ml solids. The fifth mannitol/HSA solution was used as a control with no virus. Two of the virus-containing samples were diluted with de-ionized water to about 9 mg/ml solids.

Detailed Description Text (121):

(ii) Two formulations of 6.3 ml glycine/HSA (I) in PB plus 0.4 ml adenovirus solution were prepared (29 mg/ml solids, 6.3.times.10.sup.7 iu/ml). One of the formulations was diluted with de-ionized water to 9 mg/ml solids.

Detailed Description Text (124):

Four formulations were prepared, two contained 25 ml of glycine/HSA (II) in PB plus 0.4 ml of adenovirus solution (10.5 mg/ml, 1.6.times.10.sup.7 iu/ml) and the other two contained 25 ml of glycine/HSA (II) in water plus 0.4 ml of adenovirus solution (8.6 mg/ml, 1.6.times.10.sup.7 iu/ml). The adenovirus solution underwent only one freeze/thaw cycle before usage in the above preparations. It was prepared around 10 weeks ago and was stored frozen at -70.degree. C.

Detailed Description Paragraph Table (4):

TABLE 3 Characterization of Set One Powders: Glycine/HSA in PB adenovirus formulations. Formula Dipersi. HORIBA Cascade impactor % infectivity (mg/ml) (% RSD) MMD MMAD % < 5 .mu.m retained 29 40 (25) 2.6 2.8 70 14 9 51 (1) 2.3 1.8 80 7

Detailed Description Paragraph Table (5):

TABLE 3 Characterization of Set One Powders: Glycine/HSA in PB adenovirus formulations. Formula Dipersi. HORIBA Cascade impactor % infectivity (mg/ml) (% RSD) MMD MMAD % < 5 .mu.m retained 29 40 (25) 2.6 2.8 70 14 9 51 (1) 2.3 1.8 80 7

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Term:	<div style="border: 1px solid black; padding: 2px;"> L7 same l4 </div>
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DATE: Monday, February 16, 2004 [Printable Copy](#) [Create Case](#)

<u>Set Name</u> side by side	<u>Query</u>	<u>Hit Count</u>	<u>Set Name</u> result set
<i>DB=PGPB,USPT,USOC,EPAB,JPAB,DWPI; PLUR=YES; OP=ADJ</i>			
<u>L8</u>	L7 same l4	15	<u>L8</u>
<u>L7</u>	L6 same l2	44	<u>L7</u>
<u>L6</u>	human serum albumin	10793	<u>L6</u>
<u>L5</u>	L4 same l3	7	<u>L5</u>
<u>L4</u>	concentration or titer	1296548	<u>L4</u>
<u>L3</u>	L2 with l1	60	<u>L3</u>
<u>L2</u>	adenovir\$	29148	<u>L2</u>
<u>L1</u>	HSa or human serum albumin	13009	<u>L1</u>

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L11: Entry 1 of 3

File: PGPB

May 9, 2002

DOCUMENT-IDENTIFIER: US 20020055721 A1

TITLE: Biocompatible pharmaceutical articles

Detail Description Paragraph:

[0074] Other forms of surface treatment include treating the incompatible pharmaceutical article component with solutions or suspensions containing one or more of the following: lipids and liposomes; emulsifying agents and detergents such as glycerin, sodium lauryl sulfate, sodium oleate; proteins, such as albumin, particularly human serum albumin (HSA) and bovine serum albumin (BSA); other natural polymers such as hyaluronic acid, laminin, fibronectin, fibrin, and collagen, as well as glucans and glycosaminoglycans, such as dextrans, dextran sulfate and heparin; synthetic polymers such as polyethylene glycol, polyethylene oxide, polyvinyl pyrrolidone, poloxamers, polyethylenimine, protamine sulfate, polyamidoamine dendrimers, amphiphilic peptides, RGD-oligolysine peptides, and fluorocarbons such as polytetrafluoroethylene (further synthetic polymers are listed below); contrast agents such as iohexol, blood or serum (e.g., from a patient or donor), and so forth. Treatment may be carried out by contacting the agents mentioned above with the incompatible pharmaceutical article component before that component is brought into contact with the therapeutic agent. Treatment may also be carried out by formulating the agents mentioned above directly into the solution or suspension containing the therapeutic agent. For instance, human serum albumin may be formulated into a viral suspension, such as an adenoviral suspension, in order to exert a protective or stabilizing effect. Additionally, the surface treatment may concurrently involve a cleaning process and/or sterilization process to remove surface contaminants or impurities.

INTERNATIONAL PRELIMINARY EXAMINATION REPORT

Form PCT/IPEA/409 (Box V) (July 1998)
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International application No.

PCT/US99/18515

V. Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement**1. statement**

Novelty (N)	Claims <u>1-33</u>	YES
	Claims <u>NONE</u>	NO
Inventive Step (IS)	Claims <u>1-33</u>	YES
	Claims <u>NONE</u>	NO
Industrial Applicability (IA)	Claims <u>1-33</u>	YES
	Claims <u>NONE</u>	NO

2. citations and explanations (Rule 70.7)

Claims 1-33 meet the criteria set out in PCT Article 33(2)-(4), because the prior art does not teach or fairly suggest a composition comprising a recombinant adenovirus vector and a concentration of human serum albumin (HSA) effective to stabilize the adenovirus vector at a temperature above the freezing point of water or to enhance a titer of the adenovirus vector compared to a titer in the absence of HSA, or both, in an aqueous buffer.

Claims 1-33 meet the industrial applicability as defined by PCT Article 33(4).

----- NEW CITATIONS -----
NONE